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# Marker-dependent associations among oxidative stress, growth and survival during early life in a wild mammal

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## Abstract

Oxidative stress is hypothesised to be a key physiological mechanism mediating life-history trade-offs, but evidence from wild populations experiencing natural environmental variation is limited. We tested the hypotheses that increased early-life growth rate increases oxidative stress, and that increased oxidative stress reduces first-winter survival, in wild Soay sheep (*Ovis aries*) lambs. We measured growth rate and first-winter survival for four consecutive cohorts, and measured two markers of oxidative damage (malondialdehyde, protein carbonyls) and two markers of antioxidant protection (total antioxidant capacity, superoxide dismutase) from blood samples. Faster lamb growth was weakly associated with increased malondialdehyde, but not associated with variation in the other three markers. Lambs with higher superoxide dismutase activity were more likely to survive their first winter, as were male but not female lambs with lower protein carbonyl concentrations. Survival did not vary with malondialdehyde or total antioxidant capacity. Key predictions relating oxidative stress to growth and survival were therefore supported in some oxidative stress markers but not others. This suggests that different markers capture different aspects of the complex relationships between individual oxidative state, physiology and fitness, and that overarching hypotheses relating oxidative stress to life-history variation cannot be supported or refuted by studying individual markers.

**Keywords:** Antioxidants; early-life fitness; life-history trade-offs; oxidative damage; plasma; Soay sheep.

## 1. Introduction

Oxidative stress (OS) has been proposed as a potential mediator of life-history trade-offs, providing a mechanistic explanation for variation in individual resource partitioning between self-maintenance, reproduction and growth [1,2]. Reactive oxygen species (ROS) are primarily by-products of normal aerobic metabolism [3]. Although ROS play crucial roles in cell-signalling and immune function [4], excess ROS can cause damage to macromolecules, such as lipids, proteins and DNA, and disrupt normal cellular function [5–7]. The state of imbalance between the dietary and endogenous antioxidant (AOX) defences and excess ROS, in favour of the latter, is known as OS [8].

Investment in rapid growth during early-life could confer substantial individual fitness benefits, such as predator avoidance, improved competitive ability, enhanced thermoregulation, and hence increased survival probability [9]. However, growth is highly physiologically demanding, and rapid growth may be accompanied by increased metabolic rate and thus oxygen consumption. Therefore, individuals that grow faster may be exposed to relatively more ROS [6,10 although see 11], and increased investment in growth may limit resources available for AOX defences [12]. Consequently, faster growing individuals are predicted to experience greater OS, which may ultimately impair physiological function and reduce survival probability and fitness [1]. OS could therefore provide a causal explanation for covariation in growth rates and survival, thereby shaping resource allocation, and ultimately life-history evolution. However, to date, few studies have simultaneously tested the two hypotheses that faster growth during early life is associated with OS, and that OS is associated with subsequent survival, within wild populations experiencing natural environmental variation that might constrain growth and survival. Further, despite considerable research, the degree to which OS might generally influence early-life growth and survival, or *vice versa*, remains unclear.

Some previous studies spanning captive [13–16] and wild [10,17–19] vertebrates and invertebrates found no correlations between growth rate and OS markers [20], while other studies found positive correlations between growth and oxidative damage [10,13,16–18,21,22]. There is also evidence for both negative [14] and positive correlations between growth and AOX [15,19,21]. Similarly, no overarching consensus regarding links between

OS and survival has yet emerged (summarised by [23]). OS predicted survival in some systems (e.g.[24]), but not in others (e.g. [23,25]). Further, most such studies have been conducted in captive populations under laboratory conditions. Laboratory studies allow environmental and genetic variation to be tightly controlled, and can experimentally manipulate growth or OS and sample and analyse OS across multiple occasions and tissues to examine whether effects are repeatable and tissue specific or organism-wide [26]. However, both the degree of OS experienced and its fitness consequences are likely to be highly dependent on environmental conditions. Natural populations typically experience more variable and physiologically challenging environments than laboratory populations, and might consequently show different relationships between growth, OS and survival. Therefore, although wild population studies may not readily allow experimentation or repeated sampling, they form a crucial part of our overall endeavour to understand the evolutionary causes and consequence of variation in OS [2]. Further, since environmental conditions and associated constraints on life-history allocation commonly vary among seasons and years, datasets quantifying variation in growth, OS and survival across multiple different years are ideally required to fully understand the role of physiological trade-offs in shaping life-history evolution. However, multi-year studies investigating the associations between growth and OS, and OS and survival in wild populations remain rare.

We used life-history data and plasma samples collected during four consecutive summers from a sexually dimorphic wild Soay sheep population inhabiting a naturally variable environment to quantify associations between four commonly applied markers of OS and early-life growth and survival. Specifically we used OS marker data collected on lambs to test the hypotheses that: (i) increased growth rate during early-life carries an oxidative cost and is therefore positively correlated with oxidative damage and negatively correlated with AOX; and (ii) independent of growth rate, increased oxidative damage and low levels of AOX protection reduce a lamb's first-winter survival probability. We tested whether associations between OS, growth and survival were consistent across different markers, across females and males, and across four years spanning different environmental conditions. We thereby quantify the magnitude and heterogeneity of associations between major physiological and life-history traits that could underlie key life-history trade-offs.

## **2. Materials & methods**

### **STUDY SYSTEM AND SAMPLE COLLECTION**

Plasma samples were collected from Soay sheep, resident in Village Bay, Hirta, in the St. Kilda archipelago (57°49'N 8°34'W). Since 1985, the population has been monitored intensively. Pregnant ewes are monitored closely prior to and during lambing. Lambs are caught, uniquely ear-tagged and weighed a few days after birth, between mid-April and early-May. Each lamb's sex is determined by visual examination; a scrotum sack is visible in males. The identity of each lamb's mother, and the lamb's twin status (i.e. twin or singleton), is recorded. Lambing is followed by an annual August catch where sheep are rounded up and caught in temporary traps. Individuals are identified from their tag, weighed, measured and blood sampled. Regular censuses throughout the year and mortality searches through winter and spring mean that the vast majority of carcasses are found and accurate survival data is available [27].

We used blood samples collected from lambs during four consecutive Augusts (2010-2013), to quantify OS. Blood was collected in 9ml lithium heparin vacutainers by venepuncture and stored at 4°C until processing the following day. Whole blood was spun for 10 minutes at 1008g, and the plasma supernatant was collected and stored at -80°C until further use. Samples were transported from St Kilda using either liquid nitrogen vapour shippers (2010 and 2011) or a portable -80°C freezer (2012 and 2013: Stirling Shuttle Ultra Low Portable Freezer. Triple Red Laboratory Technology, Bucks, England).

## MEASURES OF OXIDATIVE STRESS

We measured two commonly applied markers of oxidative damage: protein carbonyls (PC), a measure of protein damage, and malondialdehyde (MDA), a measure of lipid peroxidation. We also measured two markers of AOX protection: superoxide dismutase (SOD), an endogenously produced AOX enzyme, and total antioxidant capacity (TAC), a combined measure of several AOXs, both endogenous and dietary-derived. Full assay protocols are provided in Appendix S1 (see also [28]). In brief, plasma PC content (nmol/ mg protein) was quantified using a Cayman Protein Carbonyl Assay Kit (ID:10005020; Cayman Chemical Company, Ann Arbor, Michigan, USA). Protein content was quantified using the Bradford method [29]. Plasma MDA ( $\mu\text{M/L}$ ) was quantified using high-performance liquid chromatography (HPLC), following [17]. Total plasma SOD activity (U/mL) was quantified using Cayman's Superoxide Dismutase Assay Kit (ID: 706002; Cayman Chemical Company) following the manufacturer's protocol. Plasma TAC levels (mM) were estimated using Cayman's Antioxidant Assay Kit (ID: 709001; Cayman Chemical Company). These standard

assays return high within-sample repeatabilities and low inter-plate coefficients of variation [28]. The four markers were uncorrelated within years across individuals included in current analysis (Table S1, see also [28]).

We also previously measured MDA from plasma samples collected in August 2007, using identical methods [17]. This initial single-year single-marker study showed a positive association between MDA and lamb growth rate, indicating an oxidative damage cost of fast growth [17]. It has recently been suggested that MDA levels are positively related to circulating triglyceride levels in birds [30], but due to the uncertain causes of this relationship we did not attempt to correct for plasma lipid content in this study.

## STATISTICAL ANALYSIS

To test the hypothesis that rapid growth increases OS, we used 236 lambs for which we had data on all four OS markers, sex, twin status, April (birth) weight, August weight and maternal identity (2010  $n = 33$ , 18♀ & 15♂; 2011  $n = 77$ , 37♀ & 40♂; 2012  $n = 43$ , 24♀ & 19♂ and 2013  $n = 86$ , 41♀ & 45♂). A lamb's growth rate over approximately its first four months of life was calculated by dividing the difference in lamb mass at August capture and April capture by the number of days elapsed between the two capture events (following [17]. Note also that August weight and growth rate were strongly correlated in this data set; Pearson product moment correlation,  $p < 0.01$ ,  $r = 0.97$ ,  $DF = 234$ ). We fitted separate linear mixed models (LMMs) with each OS marker as the dependent variable. We fitted main fixed effects of year (4 level factor) and an individual lamb's sex (2 level factor), twin status (2 level factor) and growth rate to assess if these explained variation in marker values. Since year, sex and twin status can all influence growth rate (e.g. [17]), first order interactions between growth rate and year, growth rate and sex and growth rate and twin status were also modelled. We also included random maternal identity effects to account for non-independence of multiple lambs from the same mother. Since we also had MDA and growth data from 84 lambs sampled and measured in August 2007 [17], we refitted the LMM for MDA including these data.

To test the hypothesis that OS predicts first-winter survival, we restricted statistical analyses to samples from lambs born in 2010 and 2013 when overall first-winter mortality was 65.9% and 72.3% respectively. Samples from lambs born in 2011 and 2012 had to be excluded because first-winter mortality was very high (93.8%) and very low (4.0%) in these years respectively, meaning there was very little variation in survival to explain (see Discussion).

We included 135 lambs for which we had data on all four markers of OS, first-winter survival, August weight, sex and maternal identity (2010 n= 41, 25♀ & 16♂ and 2013 n= 94, 43♀ & 51♂). We fitted a generalised linear mixed model (GLMM) with survival as a binary dependent variable (0 = died, 1 = survived) with binomial error structure and logit link. We fitted all four OS markers as covariates within a single model, along with main effects of year (2 level factor) and sex (2 level factor), and first order interactions between each marker and year and sex. We also fitted August weight as a covariate, and fitted random maternal identity effects.

Fixed effects in LMMs and GLMMs fitted with maximum likelihood were simplified following a backwards elimination approach, using likelihood ratio tests. Terms with the lowest marginal chi-square ( $X^2$ ) statistic were sequentially dropped until only significant ( $p < 0.05$ ) explanatory variables remained. Main effects underlying retained interaction terms were also retained. All analyses were run in R version 2.15.2 [31].

### 3. Results

#### OXIDATIVE STRESS AND GROWTH RATE

We found a marginally non-significant positive association between MDA and lamb growth rate across lambs sampled during 2010-2013, but no indication of interactions between growth and year, sex or twin status (Tables 1 & S2, Figure 1). When we refitted our model including the additional MDA data from 2007, we estimated similar effect sizes but the overall effect of growth rate on MDA was marginally significant (estimated slopes of  $2.62 \pm 1.49\text{SE}$  excluding 2007 and  $2.58 \pm 1.17\text{SE}$  including 2007, Table S3). Overall, these models suggest that, as predicted, there is a positive association between MDA and growth rate, but that this association is weak and of marginal statistical significance (Figure 1, Tables 1 & S3). Our models for PC, TAC and SOD showed no significant associations with growth rate, or interactions between growth rate and year, sex or twin status (Tables 1 & S2, Figure 1). Furthermore, none of the four markers differed significantly between male and female lambs, or between singletons versus twin lambs (Table 1). However, PC, MDA and SOD, but not TAC, varied significantly among years (Table 1, Figure S1). PC concentration was highest in 2012, MDA content was lower in 2012 and 2013 than in 2010 and 2011 (and also low in 2007), and SOD activity was lowest in 2010 and highest in 2011 (Figure S1).

## OXIDATIVE STRESS AND SURVIVAL

The final model for lamb first-winter survival included main effects of SOD and August weight, and a PC-by-sex interaction (Table 2). Lambs with higher SOD activity in August were more likely to survive their first winter (Table 2, Figure 2C). Independent of this association, male lambs with high PC concentration were less likely to survive their first winter than male lambs with low PC concentrations, whereas the probability of female lamb survival did not vary with PC concentration (Table 2, Figure 2A). The PC-by-sex interaction remained significant when two male lambs with particularly high PC concentrations were excluded from the analysis ( $DF=1$ ,  $X^2=5.44$ ,  $p=0.02$ ,  $n=133$ ). MDA and TAC did not predict survival either as main effects or through interactions with year or sex (Table 2 & S4). As expected, lambs that were heavier in August were more likely to survive their first winter (Table 2).

## 4. Discussion

We tested whether relationships consistent with trade-offs between oxidative stress (OS) and growth rate were evident in Soay lambs, and tested whether OS predicted first-winter survival, a key fitness component. Moreover, we tested whether relationships were consistent across four OS markers encapsulating oxidative damage and antioxidant (AOX) protection, across four different years, and across females and males. We found some evidence of the predicted positive association between MDA and growth rate, but the estimated effect was only marginally statistically significant. The other three markers, PC, SOD and TAC, did not vary significantly with growth rate. Lamb first-winter survival showed the predicted positive association with SOD, and the predicted negative association with PC in male lambs but not in females, independent of August weight. However, survival did not vary with MDA or TAC. Therefore, overall, our results provide only partial and somewhat inconsistent support for the key hypotheses that rapid growth increases OS, and that increased OS reduces survival probability.

Some previous studies in wild birds reported the predicted negative associations between AOX levels and growth [10,18,32], but a recent meta-analysis of both wild and captive studies found evidence for an oxidative cost of increased growth only in markers of oxidative damage and not in measures of AOX [33]. The authors suggested this could reflect the inherent complexity of the AOX response to OS, with antioxidants potentially being



differentially upregulated and depleted through utilisation during growth [33]. Although our study adds to the growing literature documenting positive associations between markers of oxidative damage and growth [17,18,21], it is perhaps surprising that we only found a weak association between growth and MDA, and no association with PC. However, a study of captive domestic lambs (breed: Ile de France x INRA 401) also found no association between experimentally-induced compensatory growth and PC [34]. Surprisingly, since growth rate and August weight are both strong predictors of lamb survival on St Kilda [35], PC but not MDA predicted lamb over-winter survival. This suggests that the high MDA levels associated with faster growth have little impact on subsequent survival, and that variation in PC reflects a different aspect of oxidative state that affects survival, at least in males. Overall, therefore, our results highlight the value of relating OS to both growth and fitness components (such as survival) in natural populations; solely relating OS to growth would have led to incorrect inferences regarding the fitness consequences of variation in different markers of oxidative damage.

Indeed, our study represents a rare test of associations between OS and survival in a wild mammal. We observed substantial among-year variation in three of our four OS markers, and in lamb first-winter survival. Indeed, in 2011 and 2012 lamb mortality was so high and low, respectively, that within-year variation could not be analysed. In these years it is self-evident that among-lamb variation in OS does not drive variation in first-winter survival. This is not surprising, as extremely high or low first-winter survival on St Kilda is known to be largely driven by extrinsic factors including population density, winter climate and gastro-intestinal nematode parasite burdens [27]. Further, the high and low mortality winters were not associated with extreme mean oxidative damage or AOX values (Figure S1). For instance, mean PC was highest in 2012 when lamb mortality was low (4.0%) and mean SOD was highest in 2011 when lamb mortality was very high (93.8%). However, associations between lamb PC and SOD and first-winter survival within the two moderate mortality years (2010 and 2013) suggests that these markers may reflect aspects of the lambs' physiological state which are associated with their ability to survive winters when malnutrition, winter weather and parasite pressures are not too severe.

Our finding that lambs with high SOD were more likely to survive their first winter in moderate mortality years matches the prediction that higher investment in AOX should protect the individual from damage and promote physiological function and survival.

However, it raises the question of why SOD matches the prediction while the other markers do not. Whereas our TAC assay measures the combined effect of a suite of endogenous and exogenous AOX, our SOD assay measures a specific group of endogenously produced AOX. Variation in SOD levels in blood or tissue might therefore be a more repeatable measure of an individual's intrinsic ability to resist oxidative stress, and may be more likely to reflect the oxidative state of a lamb several months later at the onset of winter. Most other studies examining associations between OS markers (measured in blood) and survival in wild populations come from birds, and findings are quite mixed [23]. For example, low plasma MDA was associated with higher adult recruitment in European shag (*Phalacrocorax aristotelis*) nestlings [36]. King penguin chicks that did not survive the initial growth period had the lowest plasma AOX capacities and highest oxidative damage [10], although this study did not directly analyse effects of OS on survival. In one population of great tits (*Parus major*), SOD activity measured in RBCs was not associated with nestling survival [37], whilst in another population individuals that expressed intermediate activity of an important AOX enzyme, glutathione peroxidase, had highest survival probability [38]. In one of few other studies of wild mammals, MDA, but not PC or SOD, was associated with survival in adult wild mongoose (*Mungos mungo*) [39].

Further, associations between PC and survival were sex-specific in Soay lambs; although mean PC values did not differ between females and males, high PC was associated with reduced first winter survival probability in males but not females. Some other studies have reported sex-specific marker values, and/or sex-specific associations with survival. In rats (*Rattus norvegicus*), protein hydroperoxide, a cause of protein oxidation, was higher in males than in same-age females, but PC content did not differ significantly [40]. Furthermore, a sex-dependent association between OS and survival was found in alpine swifts (*Apus melba*), where RBC resistance to oxidation was higher in adult males but not females that returned the following year [41]. Soay sheep are highly polygynous and sexually dimorphic. Male lambs, under the influence of androgens, undergo more rapid growth than females, develop secondary sexual characters and are sexually mature and capable of siring offspring (although they rarely do) in the November rut in their birth year [42]. Male lambs are also less likely to survive their first winter than female lambs, at least partly due to their investment in growth and sexual development [27]. Such early-life investment in growth and reproduction might be reflected in PC values, because of the associated increase in ROS production and protein damage. This variation may predict survival in males but not females because males pay

higher survival costs of growth and reproduction in their first year. Indeed, a review of human and rat studies investigating sex differences in OS found that regardless of whether male or female rats have higher OS levels in various tissues, males are ultimately more prone to OS modulated blood pressure effects than females [43]. Thus, males might be overall more sensitive to OS than females, possibly due to the AOX effects of oestrogen [44]. Although speculative, this hypothesis could be tested by quantifying associations between PC and reproductive investment traits, and thereby testing whether oxidative costs of reproduction are sex-dependent.

Overall, our current study, alongside other studies on wild vertebrates, suggests that associations between OS markers, growth and survival are highly marker- and context-dependent. A fundamental challenge facing evolutionary and physiological ecologists aiming to understand the role of OS in mediating life-history trade-offs is therefore to understand which OS markers behave as currently predicted by life-history theory and when they do so, and which markers do not. Possible explanation are that physiological processes associated with growth, reproduction and survival elicit certain types of oxidative damage and AOX but not others (see [39]), or that different types of OS are differentially affected by independent biotic or abiotic factors that also affect growth and survival. Another possibility results from the fact that most wild population studies, including ours, are constrained to sample blood, and to sample individuals at single logistically-feasible time points that may not coincide with expression of key life-history traits. Testing relationships between marker values and life-history traits therefore requires that blood-based OS markers reflect organism-wide oxidative state, and that individual measures are repeatable over time. However, some laboratory studies suggest that concentrations of particular OS markers may be only weakly correlated across tissues [43,45,46] and that some OS markers show circadian variation [47]. Yet, to date, studies that considered multiple tissues have all been cross-sectional, and longitudinal studies of OS markers measured in blood rarely report the within-individual repeatability (although see [48]). Longitudinal field- and laboratory studies that quantify the repeatability of different OS markers across tissues- and individuals over different times and life-history stages are challenging to implement, but are urgently required if we are to understand the role of OS in mediating life-history evolution.

**Data accessibility.** Data will be made available on the Dryad Digital Repository, following publication.

**Author's contributions.** LLC, DNH, JMR and CS designed the study and analyses. Data were collected by LLC, JGP, KAW, JMP and DNH. LLC carried out laboratory and statistical analyses and drafted the manuscript. CS, JDB and DNH supplied laboratory space, equipment and reagents. The manuscript was written by LLC, DNH and JMR, with editorial input from all other authors. All authors gave approval for publication.

**Competing interests.** None declared.

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## References

1. Monaghan, P., Metcalfe, N. B. & Torres, R. 2009 Oxidative stress as a mediator of life history trade-offs: mechanisms, measurements and interpretation. *Ecol. Lett.* **12**, 75–92. (doi:10.1111/j.1461-0248.2008.01258.x)
2. Selman, C., Blount, J. D., Nussey, D. H. & Speakman, J. R. 2012 Oxidative damage, ageing, and life-history evolution: where now? *Trends Ecol. Evol.* **27**, 570–577. (doi:10.1016/j.tree.2012.06.006)
3. Dowling, D. K. & Simmons, L. W. 2009 Reactive oxygen species as universal constraints in life-history evolution. *Proc. R. Soc. B* **276**, 1737–1745. (doi:10.1098/rspb.2008.1791)
4. von Schantz, T., Bensch, S., Grahn, M., Hasselquist, D. & Wittzell, H. 1999 Good genes, oxidative stress and condition-dependent sexual signals. *Proc. R. Soc. London B* **266**, 1–12. (doi:10.1098/rspb.1999.0597)
5. Harman, D. 1956 Aging: a theory based on free radical and radiation chemistry. *J. Gerontol.* **2**, 298–300.
6. Beckman, K. B. & Ames, B. N. 1998 The free radical theory of aging matures. *Physiol. Rev.* **78**, 547–581.
7. Buffenstein, R., Edrey, Y. H., Yang, T. & Mele, J. 2008 The oxidative stress theory of aging: embattled or invincible? Insights from non-traditional model organisms. *Age (Omaha)*. **30**, 99–109. (doi:10.1007/s11357-008-9058-z)
8. Sies, H. & Jones, D. 2007 Oxidative stress. In *Encyclopedia of Stress*, pp. 45–48.
9. Arnott, S. A., Chiba, S. & Conover, D. O. 2006 Evolution of intrinsic growth rate: metabolic costs drive trade-offs between growth and swimming performance in *Menidia menidia*. *Evolution (N. Y.)*. **60**, 1269–1278. (doi:10.1111/j.0014-

- 3820.2006.tb01204.x)
10. Geiger, S., Le Vaillant, M., Lebard, T., Reichert, S., Stier, A., LE Maho, Y. & Criscuolo, F. 2012 Catching-up but telomere loss: half-opening the black box of growth and ageing trade-off in wild king penguin chicks. *Mol. Ecol.* **21**, 1500–1510. (doi:10.1111/j.1365-294X.2011.05331.x)
  11. Salin, K. et al. 2015 Individuals with higher metabolic rates have lower levels of reactive oxygen species in vivo. *Biol. Lett.* **11**, 4–7. (doi:10.1098/rsbl.2015.0538)
  12. Dmitriew, C. M. 2011 The evolution of growth trajectories: what limits growth rate? *Biol. Rev.* **86**, 97–116. (doi:10.1111/j.1469-185X.2010.00136.x)
  13. Rollo, C. D., Carlson, J. & Sawada, M. 1996 Accelerated aging of giant transgenic mice is associated with elevated free radical processes. *Can. J. Zool.* , 606–620.
  14. Alonso-Alvarez, C., Bertrand, S., Faivre, B. & Sorci, G. 2007 Increased susceptibility to oxidative damage as a cost of accelerated somatic growth in zebra finches. *Funct. Ecol.* **21**, 873–879. (doi:10.1111/j.1365-2435.2007.01300.x)
  15. Almaida-Pagan, P. F., Lucas-Sanchez, A. & Tocher, D. R. 2014 Changes in mitochondrial membrane composition and oxidative status during rapid growth, maturation and aging in zebrafish, *Danio rerio*. *Biochim. Biophys. Acta - Mol. Cell Biol. Lipids* **1841**, 1003–1011. (doi:10.1016/j.bbalip.2014.04.004)
  16. Tsunekage, T. & Ricklefs, R. E. 2015 Increased lipid peroxidation occurs during development in Japanese quail (*Coturnix japonica*) embryos. *Br. Poult. Sci.* **56**, 262–266. (doi:10.1080/00071668.2014.994592)
  17. Nussey, D. H., Pemberton, J. M., Pilkington, J. G. & Blount, J. D. 2009 Life history correlates of oxidative damage in a free-living mammal population. *Funct. Ecol.* **23**, 809–817. (doi:10.1111/j.1365-2435.2009.01555.x)
  18. Stier, A., Delestrade, A., Zahn, S., Arrivé, M., Criscuolo, F. & Massemin-Challet, S. 2014 Elevation impacts the balance between growth and oxidative stress in coal tits. *Oecologia* **175**, 791–800. (doi:10.1007/s00442-014-2946-2)
  19. Stier, A., Massemin, S., Zahn, S., Tissier, M. L. & Criscuolo, F. 2015 Starting with a handicap: effects of asynchronous hatching on growth rate, oxidative stress and telomere dynamics in free-living great tits. *Oecologia* **179**, 999–1010. (doi:10.1007/s00442-015-3429-9)
  20. Rosa, C. E., Figueiredo, M. A., Lanes, C. F. C., Almeida, D. V, Monserrat, J. M. & Marins, L. F. 2008 Metabolic rate and reactive oxygen species production in different genotypes of GH-transgenic zebrafish. *Comp. Biochem. Physiol. Part B* **149**, 209–214.

(doi:10.1016/j.cbpb.2007.09.010)

21. Almroth, B. C., Johnsson, J. I., Devlin, R. & Sturve, J. 2012 Oxidative stress in growth hormone transgenic coho salmon with compressed lifespan - a model for addressing aging. *Free Radic. Res.* **46**, 1183–1189. (doi:10.3109/10715762.2012.698009)
22. Guerra, C., Zenteno-Savín, T., Maeda-Martínez, A. N., Philipp, E. E. R. & Abele, D. 2012 Changes in oxidative stress parameters in relation to age, growth and reproduction in the short-lived catarina scallop *Argopecten ventricosus* reared in its natural environment. *Comp. Biochem. Physiol. Part A.* **162**, 421–430. (doi:10.1016/j.cbpa.2012.04.018)
23. Costantini, D. 2014 *Oxidative Stress and Hormesis in Evolutionary Ecology and Physiology*. (doi:10.1007/978-3-642-54663-1)
24. Freeman-Gallant, C. R., Amidon, J., Berdy, B., Wein, S., Taff, C. C. & Haussmann, M. F. 2011 Oxidative damage to DNA related to survivorship and carotenoid-based sexual ornamentation in the common yellowthroat. *Biol. Lett.* **7**, 429–432. (doi:10.1098/rsbl.2010.1186)
25. De Coster, G., De Neve, L., Verhulst, S. & Lens, L. 2012 Maternal effects reduce oxidative stress in female nestlings under high parasite load. *J. Avian Biol.* **43**, 177–185. (doi:10.1111/j.1600-048X.2012.05551.x)
26. Speakman, J. R. et al. 2015 Oxidative stress and life histories: unresolved issues and current needs. *Ecol. Evol.* **5**, 5745–5757. (doi:10.1002/ece3.1790)
27. Clutton-Brock, T. H. & Pemberton, J. M. 2004 *Soay Sheep: Dynamics & Selection in an Island Population*. Cambridge: Cambridge University Press.
28. Christensen, L. L., Selman, C., Blount, J. D., Pilkington, J. G., Watt, K. A., Pemberton, J. M., Reid, J. M. & Nussey, D. H. 2015 Plasma markers of oxidative stress are uncorrelated in a wild mammal. *Ecol. Evol.* **5**, 5096–5108. (doi:10.1002/ece3.1771)
29. Bradford, M. M. 1976 A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal. Biochem.* **72**, 248–254.
30. Pérez-Rodríguez, L., Romero-Haro, A. A., Sternalski, A., Muriel, J., Mougeot, F., Gil, D. & Alonso-Alvarez, C. 2015 Measuring oxidative stress: the confounding effect of lipid concentration in measures of lipid peroxidation. *Physiol. Biochem. Zool.* **88**, 345–351. (doi:10.1086/680688)
31. R Core Team 2012 R: A language and environment for statistical computing. R

Foundation for Statistical Computing, Vienna, Austria.

32. Kim, S.-Y., Noguera, J. C., Morales, J. & Velando, A. 2011 Quantitative genetic evidence for trade-off between growth and resistance to oxidative stress in a wild bird. *Evol. Ecol.* **25**, 461–472. (doi:10.1007/s10682-010-9426-x)
33. Smith, S. M., Nager, R. G. & Costantini, D. 2016 Meta-analysis indicates that oxidative stress is both a constraint on and a cost of growth. *Ecol. Evol.* **6**, 2833–2842. (doi:10.1002/ece3.2080)
34. Savary-Auzeloux, I., Durand, D., Gruffat, D., Bauchart, D. & Ortigues-Marty, I. 2008 Food restriction and refeeding in lambs influence muscle antioxidant status. *Animal* **2**, 738–745. (doi:10.1017/S1751731108001742)
35. Clutton-Brock, T. H., Stevenson, I. R., Marrow, P., MacColl, a D., Houston, a I. & McNamara, J. M. 1996 Population fluctuations, reproductive costs and life-history tactics in female Soay sheep. *J. Anim. Ecol.* **65**, 675–689. (doi:10.2307/5667)
36. Noguera, J. C., Kim, S.-Y. & Velando, a. 2012 Pre-fledgling oxidative damage predicts recruitment in a long-lived bird. *Biol. Lett.* **8**, 61–63. (doi:10.1098/rsbl.2011.0756)
37. Koivula, M. J., Kanerva, M., Salminen, J.-P., Nikinmaa, M. & Eeva, T. 2011 Metal pollution indirectly increases oxidative stress in great tit (*Parus major*) nestlings. *Environ. Res.* **111**, 362–370. (doi:10.1016/j.envres.2011.01.005)
38. Norte, A. C., Ramos, J. A., Araujo, P. M., Sousa, J. P. & Sheldon, B. C. 2008 Health-state variables and enzymatic biomarkers as survival predictors in nestling great tits (*Parus Major*): effects of environmental conditions. *Auk* **125**, 943–952. (doi:10.1525/auk.2008.07188)
39. Vitikainen, E. I. K. et al. 2016 Evidence of oxidative shielding of offspring in a wild mammal. *Front. Ecol. Evol.* **4**, 1–10. (doi:10.3389/fevo.2016.00058)
40. Kayali, R., Çakatay, U. & Tekeli, F. 2007 Male rats exhibit higher oxidative protein damage than females of the same chronological age. *Mech. Ageing Dev.* **128**, 365–369. (doi:10.1016/j.mad.2007.03.003)
41. Bize, P., Devevey, G., Monaghan, P., Doligez, B. & Christe, P. 2008 Fecundity and survival in relation to resistance to oxidative stress in a free-living bird. *Ecology* **89**, 2584–2593.
42. Coltman, D. W., Smith, J. A., Bancroft, D. R., Pilkington, J., MacColl, A. D., Clutton-Brock, T. H. & Pemberton, J. M. 1999 Density-dependent variation in lifetime breeding success and natural and sexual selection in Soay rams. *Am. Nat.* **154**, 730–

746. (doi:10.1086/303274)
43. Sartori-Valinotti, J. C., Iliescu, R., Fortepiani, L. A., Yanes, L. L. & Reckelhoff, J. F. 2007 Sex differences in oxidative stress and the impact on blood pressure control and cardiovascular disease. *Clin. Exp. Pharmacol. Physiol.* **34**, 938–945. (doi:10.1111/j.1440-1681.2007.04643.x)
  44. Strehlow, K., Rotter, S., Wassmann, S., Adam, O., Grohé, C., Laufs, K., Böhm, M. & Nickenig, G. 2003 Modulation of antioxidant enzyme expression and function by estrogen. *Circ. Res.* **93**, 170–177. (doi:10.1161/01.RES.0000082334.17947.11)
  45. Argüelles, S., García, S., Maldonado, M., Machado, A. & Ayala, A. 2004 Do the serum oxidative stress biomarkers provide a reasonable index of the general oxidative stress status? *Biochim. Biophys. Acta* **1674**, 251–259. (doi:10.1016/j.bbagen.2004.06.023)
  46. Tkachenko, H., Kurhaluk, N., Grudniewska, J. & Andriichuk, A. 2014 Tissue-specific responses of oxidative stress biomarkers and antioxidant defenses in rainbow trout *Oncorhynchus mykiss* during a vaccination against furunculosis. *Fish Physiol. Biochem.* **40**, 1289–1300. (doi:10.1007/s10695-014-9924-9)
  47. Lewis, N. A., Newell, J., Burden, R., Howatson, G. & Pedlar, C. R. 2016 Critical Difference and Biological Variation in Biomarkers of Oxidative Stress and Nutritional Status in Athletes. *PLoS One* **11**, 1–12. (doi:10.1371/journal.pone.0149927)
  48. Dahwa, R., Fassett, R. G., Wang, Z., Briskey, D., Mallard, A. R. & Coombes, J. S. 2014 Variability of oxidative stress biomarkers in hemodialysis patients. *Biomarkers* **19**, 154–158. (doi:10.3109/1354750X.2013.867533)



**Table 1.** Linear mixed models of four different biomarkers of oxidative stress in response to growth rate (kg day<sup>-1</sup>) in Soay sheep lambs and associated degrees of freedom (DF), estimates and standard errors, for (A) protein carbonyls (PC), (B) malondialdehyde (MDA), (C) superoxide dismutase (SOD) and (D) total antioxidant capacity (TAC). Intercepts were set to female lambs in 2010. All models show terms retained after model simplification, along with dropped main effects in order of elimination (full models, including interactions and random effects, are shown in Table S2). All models used data from 236 lambs collected in 2010-2013.

**Table 2.** Generalized linear mixed model of first winter survival in relation to four different biomarkers of oxidative stress (protein carbonyls (PC), malondialdehyde (MDA), superoxide dismutase (SOD) and total antioxidant capacity (TAC)) in Soay sheep lambs and associated degrees of freedom (DF), estimates and standard errors. The intercept was set to female lambs in 2010. Model terms retained after simplification are shown, along with dropped main effects in order of elimination (the full model, including interactions and random effects, is shown in Table S4). The model used data from 135 lambs collected in 2010 and 2013.

**Figure 1.** Relationships between four markers of oxidative stress markers and growth rate (increase in mass over time, kg day<sup>-1</sup>) in Soay sheep lambs, with year (2010= grey solid dots, 2011= black solid dots, 2012= black open dots and 2013= grey open dots, total n= 236). (A) Protein carbonyl (PC, nmol/ mg protein), (B) malondialdehyde (MDA,  $\mu$ M/L), (C) superoxide dismutase (SOD, U/mL) and (D) total antioxidant capacity (TAC, mM). Lines show predicted marker levels by year (2010= grey line, 2011= black line, 2012= black dotted line and 2013= grey dotted line).

**Figure 2.** Relationships between observed first-winter survival (1: survived, 0: died) and predicted survival probabilities for Soay sheep lambs and (A) protein carbonyls (PC, nmol/ mg protein), (B) malondialdehyde (MDA,  $\mu$ M/L), (C) superoxide dismutase (SOD, U/mL) and (D) total antioxidant capacity (TAC, mM), respectively. Thick and thin lines show predicted survival  $\pm$  one standard error. Males (black) and females (orange) of all sampled lambs of 2010 and 2013 (n= 135).

**Table 1.**

Term	DF	X <sup>2</sup>	p-value	Fixed effects	Estimate	Standard error
<b>(A) Protein carbonyls</b>						
Final model (conditional R <sup>2</sup> = 0.51)						
Year	3	170.98	<0.001	Intercept	0.37	0.05
				2011	-0.005	0.05
				2012	0.71	0.06
				2013	0.19	0.05
Dropped terms						
Twin	1	0.51	0.47		-0.04	0.05
Growth	1	0.19	0.66		-0.40	0.92
Sex	1	0.67	0.41		0.03	0.05
<b>(B) Malondialdehyde</b>						
Final model (conditional R <sup>2</sup> = 0.44)						
Year	3	49.07	<0.001	Intercept	1.68	0.08
				2011	0.12	0.09
				2012	-0.38	0.10
				2013	-0.25	0.09
Dropped terms						
Sex	1	<0.01	0.99		<-0.01	0.06
Twin	1	0.49	0.49		-0.06	0.09
Growth	1	3.05	0.08		2.62	1.49
<b>(C) Superoxide dismutase</b>						
Final model (conditional R <sup>2</sup> = 0.64)						
Year	3	108.79	<0.001	Intercept	4.23	0.61
				2011	7.65	0.71
				2012	4.07	0.76
				2013	3.49	0.70
Dropped terms						
Growth	1	0.17	0.68		-5.73	13.91
Sex	1	0.12	0.73		0.14	0.41
Twin	1	0.55	0.46		-0.44	0.60
<b>(D) Total antioxidant capacity</b>						
Final model <i>None</i>						
Dropped terms						
Sex	1	0.03	0.87		0.03	0.16
Twin	1	0.57	0.45		-0.16	0.22
Growth	1	1.62	0.20		-4.77	3.75
Year	3	6.26	0.10			
2011					0.14	0.24
2012					0.17	0.26
2013					0.44	0.23

Table 2.

Term	DF	X <sup>2</sup>	p-value	Fixed effects	Estimate	Standard error
Survival						
Final model (conditional R <sup>2</sup> = 0.54)				Intercept	-6.82	2.63
SOD	1	4.30	0.04	SOD	0.16	0.09
August weight	1	15.32	<0.001	August weight	0.42	0.17
PC*Sex	1	5.74	0.02	PC*Sex	-6.02	3.12
Sex				Sex	1.24	1.30
PC				PC	-0.23	1.56
Dropped terms						
MDA	1	1.04	0.31		-0.56	0.52
TAC	1	0.98	0.32		0.17	0.18
Year	1	1.66	0.20		-0.83	0.70

Figure 1.

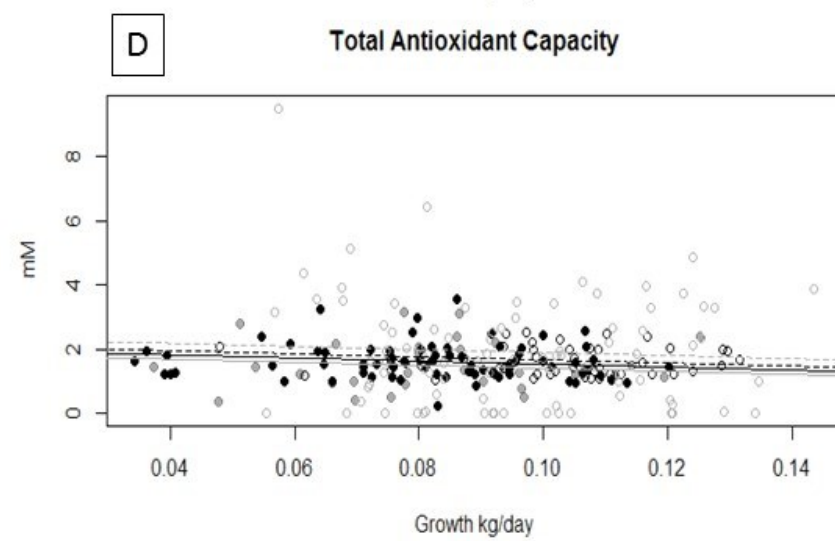
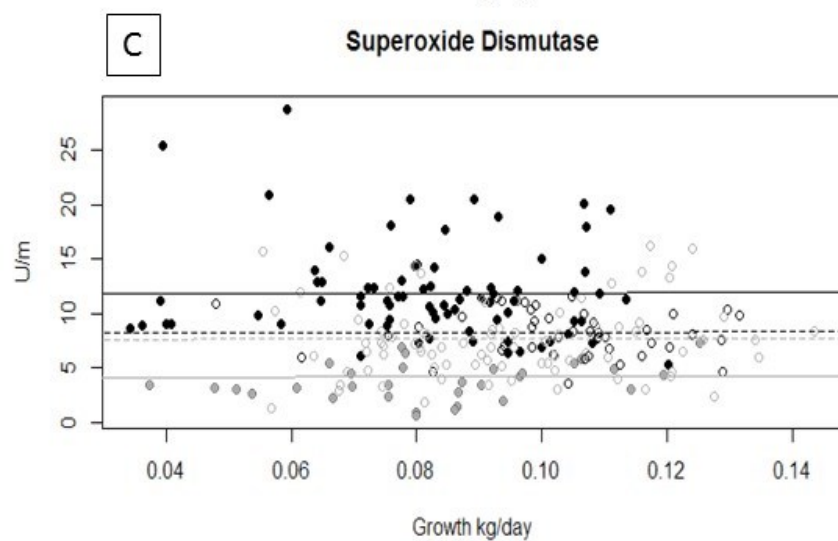
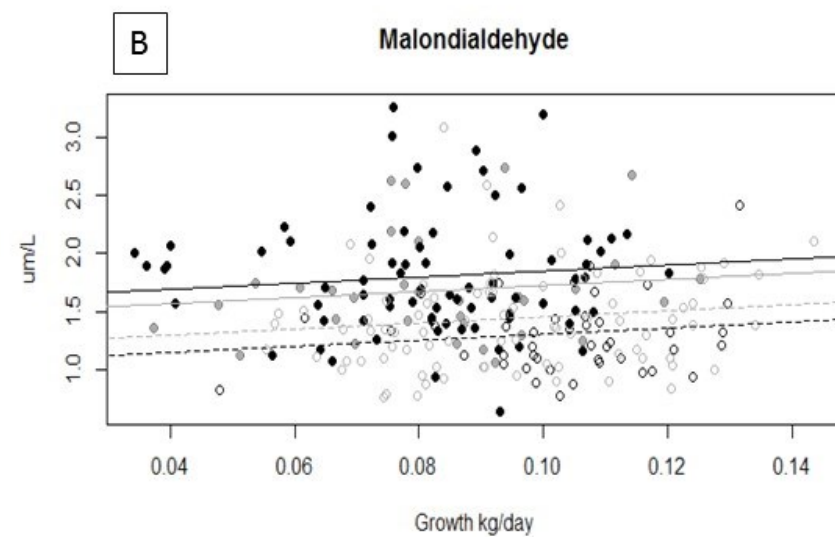
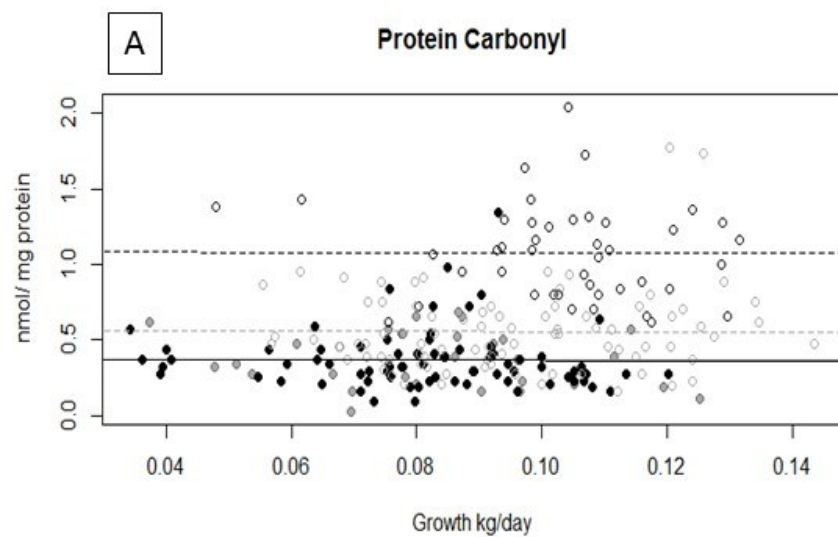


Figure 2.

